

Three New Limonoids from the Leaves of *Cipadessa cinerascens*

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Three new limonoids, cipadesins G–I (**1–3**), together with four known ones, were isolated from the leaves of *Cipadessa cinerascens*. Their structures were elucidated on the basis of 1D- and 2D-NMR data.

Introduction. – *Cipadessa cinerascens* (PELL.) HAND.–MAZZ. (Meliaceae) is a shrub mainly distributed in the Chinese southwest provinces such as Sichuan, Yunnan, Guizhou, and Guangxi [1]. Its leaves, barks, and roots have been used in Chinese folk medicine for the treatment of stomachache, dysentery, rheumatism, malaria, scald, and skin itch [2]. Previously, flavonoids and their glucosides were isolated from its leaves [3–5], and several structural novel limonoids were also reported [6–13]. This article reports the isolation and structural elucidation of the three new tetranortriterpenoids cipadesins G–I¹⁾ (**1–3**), together with cipadesin B (**4**) [6], cipadesin D (**5**) [7], cineracipadesin E (**6**) [8], and cipadesin C (**7**) [6] from the leaves of *C. cinerascens*.

Results and Discussion. – Cipadesin G (**1**) was isolated as white powder. Its molecular formula was deduced as C₂₉H₃₈O₉ from the HR-ESI-MS (m/z 553.2432 ($[M + Na]^+$)). The IR spectrum showed absorption bands for ketone C=O (1743 cm⁻¹) and ester C=O (1735 cm⁻¹) moieties. The ¹H-NMR data (Table 1) of **1** exhibited six Me *s* at δ (H) 1.04 (*s*, Me(18)), 0.91 (*s*, Me(19)), 1.05 (*s*, Me(28)), 0.86 (*s*, Me(29)), 2.06 (*s*, AcO–C(3)), and 3.71 (*s*, MeO) and a Me *d* at δ (H) 1.29 (*d*, $J = 7.0$ Hz, Me(30)), five of which were skeletal Me groups, besides three olefinic H-atoms at δ (H) 8.16 (*s*, H–C(21)), 6.66 (*s*, H–C(22)), and 7.47 (*s*, H–C(23)), suggesting the presence of a furan ring. The ¹³C-NMR and DEPT spectra (Table 2) supported the existence of a furan ring by δ (C) 121.1 (*s*, C(20)), 142.2 (*d*, C(21)), 110.5 (*d*, C(22)), and 143.5 (*d*, C(23)), and also revealed a C=O group at δ (C) 212.9 (*s*, C(9)). All of these were characteristic of a cipadesin-type limonoid, in which rings A and C were connected *via* C(10) and C(11) [6]. HMBCs of H–C(5) with C(10) and C(11), and Me(19) with C(11) supported the connection. Further detailed 2D-NMR analysis confirmed the cipadesin-type limonoid skeleton as shown in Fig. 1. An AcO group was located at C(3) by a HMBC cross-peak H–C(3)/C=O of the AcO group, and a ROESY correlation H–C(3)/Me(19) indicated β -configuration of the AcO group. Therefore, the structure of **1** was established.

¹⁾ Arbitrary atom numbering; for systematic names, see *Exper. Part*.

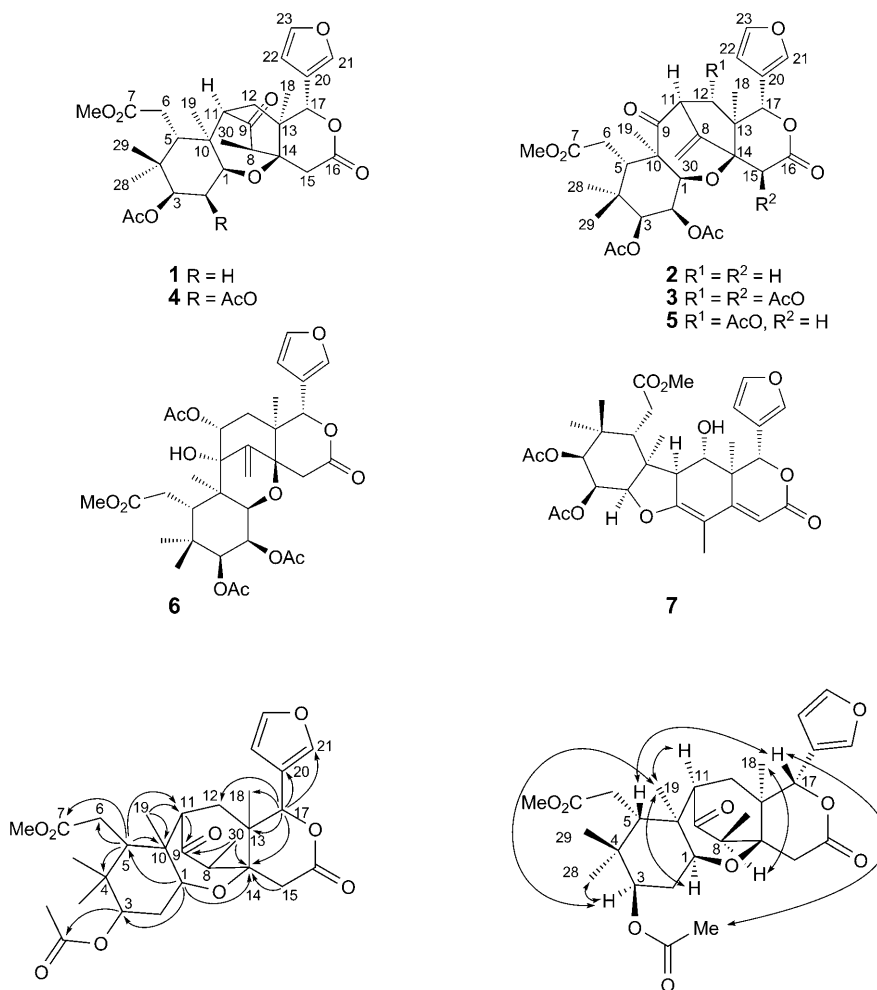


Fig. 1. Important HMBCs (H → C) and ROESY (H ↔ H) correlations of compound **1**

Cipadesin H (**2**) had a molecular formula $C_{31}H_{38}O_{11}$ as determined by the HR-ESI-MS (m/z 609.2284 ($[M + Na]^+$)). The IR peaks at 1746 and 1737 cm^{-1} suggested the presence of ketone C=O and ester C=O groups, respectively. The 1D-NMR spectra (Tables 1 and 2) showed four skeletal Me groups at $\delta(H)$ 0.87 (*s*, Me(18)), 1.09 (*s*, Me(19)), 1.05 (*s*, Me(28)), and 0.81 (*s*, Me(29)) and $\delta(C)$ 17.6 (*q*, Me(18)), 19.3 (*q*, Me(19)), 22.8 (*q*, Me(28)), and 27.5 (*q*, Me(19)), and an exocyclic C=C group at $\delta(H)$ 5.40 (*s*, H_a-C(30)) and 5.18 (*s*, H_b-C(30)) and $\delta(C)$ 114.2 (*t*, C(30)) and 143.5 (*s*, C(8)), together with a furan ring at $\delta(H)$ 7.55 (*s*, H-C(21)), 6.45 (*t*, $J=1.0$ Hz, H-C(22)), and 7.46 (*t*, $J=1.0$ Hz, H-C(23)) and $\delta(C)$ 121.8 (*s*, C(20)), 139.7 (*d*, C(21)), 108.5 (*d*, C(22)), and 144.2 (*d*, C(23)). Besides, a C=O group at $\delta(C)$ 209.2 (*s*, C(9)) was observed in the ^{13}C -NMR spectra. The above signals suggested that

Table 1. $^1\text{H-NMR}$ Data (500 MHz, CDCl_3) of Compounds **1–3**. δ in ppm, J in Hz.

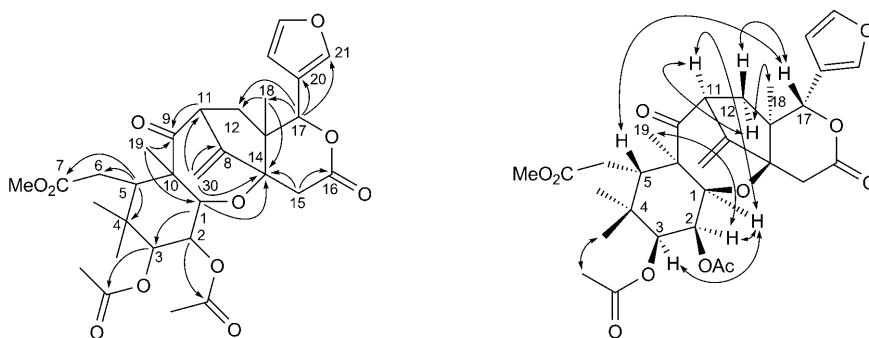
	1	2	3
$\text{H}_\alpha\text{-C}(1)$	3.40 (<i>t</i> , $J=3.0$)	4.18 (<i>d</i> , $J=2.8$)	4.33 (<i>d</i> , $J=3.4$)
$\text{CH}_2(2)$ or $\text{H}_\alpha\text{-C}(2)$	2.03–2.07 (<i>m</i> , 2 H)	5.21 (<i>t</i> , $J=2.8$)	5.32 (<i>t</i> , $J=3.4$)
$\text{H}_\alpha\text{-C}(3)$	4.71 (<i>t</i> , $J=2.5$)	5.11 (<i>d</i> , $J=2.8$)	5.11 (<i>d</i> , $J=3.4$)
$\text{H}_\alpha\text{-C}(5)$	2.79 (<i>d</i> , $J=10.0$)	2.94 (<i>t</i> , $J=4.5$)	3.08 (<i>d</i> , $J=6.0$)
$\text{H}_\alpha\text{-C}(6)$	2.05–2.07 (<i>m</i>)	2.25 (<i>dd</i> , $J=18.5, 4.5$)	2.25–2.27 (<i>m</i>)
$\text{H}_\beta\text{-C}(6)$	2.44 (<i>dd</i> , $J=17.5, 10.0$)	2.89 (<i>dd</i> , $J=18.5, 4.5$)	2.79 (<i>d</i> , $J=17.0$)
$\text{H}_\alpha\text{-C}(8)$	2.62 (<i>d</i> , $J=7.0$)	–	–
$\text{H}_\alpha\text{-C}(11)$	2.39 (<i>s</i>)	3.53 (<i>dd</i> , 11.0, 4.5)	3.36 (<i>d</i> , $J=2.5$)
$\text{H}_\alpha\text{-C}(12)$	1.25–1.30 (<i>m</i>)	1.70–1.76 (<i>m</i>)	–
$\text{H}_\beta\text{-C}(12)$	2.61–2.38 (<i>m</i>)	2.80–2.83 (<i>m</i>)	5.63 (<i>d</i> , $J=2.5$)
$\text{H}_\alpha\text{-C}(15)$	2.73 (<i>d</i> , $J=17.5$)	2.79 (<i>d</i> , $J=18.0$)	6.00 (<i>s</i>)
$\text{H}_\beta\text{-C}(15)$	2.56 (<i>d</i> , $J=17.5$)	2.83 (<i>d</i> , $J=18.0$)	–
$\text{H}_\beta\text{-C}(17)$	6.52 (<i>s</i>)	6.40 (<i>s</i>)	6.53 (<i>s</i>)
Me(18)	1.04 (<i>s</i>)	0.87 (<i>s</i>)	1.09 (<i>s</i>)
Me(19)	0.91 (<i>s</i>)	1.09 (<i>s</i>)	1.14 (<i>s</i>)
H–C(21)	8.16 (<i>s</i>)	7.55 (<i>s</i>)	7.56 (<i>s</i>)
H–C(22)	6.66 (<i>s</i>)	6.45 (<i>t</i> , $J=1.0$)	6.41 (<i>s</i>)
H–C(23)	7.47 (<i>s</i>)	7.46 (<i>t</i> , $J=1.0$)	7.43 (<i>s</i>)
Me(28)	1.05 (<i>s</i>)	1.05 (<i>s</i>)	1.07 (<i>s</i>)
Me(29)	0.86 (<i>s</i>)	0.81 (<i>s</i>)	0.85 (<i>s</i>)
Me(30) or $\text{CH}_2(30)$	1.29 (<i>d</i> , $J=7.0$)	5.40 (<i>s</i>), 5.18 (<i>s</i>)	5.35 (<i>s</i>), 5.11 (<i>s</i>)
MeO	3.71 (<i>s</i>)	3.67 (<i>s</i>)	3.64 (<i>s</i>)
AcO–C(2)	–	2.11 (<i>s</i>)	2.12 (<i>s</i>)
AcO–C(3)	2.06 (<i>s</i>)	2.08 (<i>s</i>)	2.09 (<i>s</i>)
AcO–C(12)	–	–	1.86 (<i>s</i>)
AcO–C(15)	–	–	2.22 (<i>s</i>)

compound **2** was a trijugin-type limonoid characterized by a five-membered ring *C* with an exocyclic C=O group at C(9) [14]. By comprehensive 2D-NMR analysis, the trijugin-type limonoid framework was finally determined for compound **2** as shown in Fig. 2. Two AcO groups were placed at C(2) and C(3) by the HMBC cross-peaks H–C(2)/C=O of AcO–C(2), and H–C(3)/C=O of AcO–C(3). The ROESY correlations H–C(2)/H–C(1) and Me(19) and H–C(2)/Me(28) as well as H–C(3)/Me(28) indicated the relative β -configuration of these two AcO groups. Accordingly, the structure of compound **2** was identified.

The molecular formula of cipadesin I (**3**) was determined as $\text{C}_{35}\text{H}_{42}\text{O}_{15}$ from the HR-ESI-MS (m/z 725.2442 [$M + \text{Na}]^+$). Its IR spectrum showed absorption bands similar to those of **2**. The ^1H - and ^{13}C -NMR data (Tables 1 and 2) of **3** resemble those of **2**, except for the absence of two CH_2 groups and the presence of two additional CH–O groups at $\delta(\text{H})$ 5.63 (*d*, $J=2.5$ Hz, H–C(12)) and 6.00 (*s*, H–C(15)) and $\delta(\text{C})$ 76.3 (*d*, C(12)) and 69.7 (*d*, C(15)), and two more AcO groups at $\delta(\text{H})$ 1.86 (*s*, AcO–C(12)) and 2.22 (*s*, AcO–C(15)), and $\delta(\text{C})$ 168.8 and 20.4 (*s* and *q*, AcO–C(12)), and 169.7 and 20.3 (*s* and *q*, AcO–C(15)). HMBC Cross-peaks H–C(12)/C=O of AcO–C(12), and H–C(15)/C=O of AcO–C(15) definitely positioned these two supplementary AcO groups at C(12) and C(15). The ROESY correlations H–C(12)/H–C(5) and H–C(17) and H–C(15)/H–C(1) assigned the

Table 2. ^{13}C -NMR Data (125 MHz, CDCl_3) of Compounds **1–3**. δ in ppm.

	1	2	3		1	2	3
C(1)	75.5 (<i>d</i>)	73.7 (<i>d</i>)	75.7 (<i>d</i>)	C(19)	18.9 (<i>q</i>)	19.3 (<i>q</i>)	20.0 (<i>q</i>)
C(2)	27.5 (<i>t</i>)	65.4 (<i>d</i>)	65.8 (<i>d</i>)	C(20)	121.1 (<i>s</i>)	121.8 (<i>s</i>)	120.4 (<i>s</i>)
C(3)	75.4 (<i>d</i>)	74.4 (<i>d</i>)	75.2 (<i>d</i>)	C(21)	142.2 (<i>d</i>)	139.7 (<i>d</i>)	140.7 (<i>d</i>)
C(4)	37.8 (<i>s</i>)	39.0 (<i>s</i>)	39.1 (<i>s</i>)	C(22)	110.5 (<i>d</i>)	108.5 (<i>d</i>)	108.8 (<i>d</i>)
C(5)	37.0 (<i>d</i>)	37.6 (<i>d</i>)	35.2 (<i>d</i>)	C(23)	143.5 (<i>d</i>)	144.2 (<i>d</i>)	143.4 (<i>d</i>)
C(6)	30.7 (<i>t</i>)	29.1 (<i>t</i>)	28.7 (<i>t</i>)	C(28)	21.2 (<i>q</i>)	22.8 (<i>q</i>)	22.7 (<i>q</i>)
C(7)	174.2 (<i>s</i>)	174.2 (<i>s</i>)	173.7 (<i>s</i>)	C(29)	28.0 (<i>q</i>)	27.5 (<i>q</i>)	27.3 (<i>q</i>)
C(8)	44.4 (<i>d</i>)	143.5 (<i>s</i>)	139.6 (<i>s</i>)	C(30)	10.6 (<i>q</i>)	114.2 (<i>t</i>)	116.6 (<i>t</i>)
C(9)	212.9 (<i>s</i>)	209.2 (<i>s</i>)	204.4 (<i>s</i>)	MeO	52.2 (<i>q</i>)	51.8 (<i>q</i>)	51.9 (<i>q</i>)
C(10)	44.0 (<i>s</i>)	55.9 (<i>s</i>)	56.6 (<i>s</i>)	AcO–C(2)		170.4 (<i>s</i>),	170.9 (<i>s</i>),
C(11)	57.5 (<i>d</i>)	58.5 (<i>d</i>)	69.4 (<i>d</i>)			20.8 (<i>q</i>)	21.2 (<i>q</i>)
C(12)	29.0 (<i>t</i>)	37.6 (<i>t</i>)	76.3 (<i>d</i>)	AcO–C(3)	170.9 (<i>s</i>),	170.8 (<i>s</i>),	170.8 (<i>s</i>),
C(13)	40.3 (<i>s</i>)	45.5 (<i>s</i>)	53.3 (<i>s</i>)		21.0 (<i>q</i>)	20.4 (<i>q</i>)	20.6 (<i>q</i>)
C(14)	79.9 (<i>s</i>)	87.9 (<i>s</i>)	88.9 (<i>s</i>)	AcO–C(12)			168.8 (<i>s</i>),
C(15)	38.7 (<i>t</i>)	33.5 (<i>t</i>)	69.7 (<i>d</i>)				20.4 (<i>q</i>)
C(16)	169.0 (<i>s</i>)	168.7 (<i>s</i>)	166.4 (<i>s</i>)	AcO–C(15)			169.7 (<i>s</i>),
C(17)	79.6 (<i>d</i>)	79.4 (<i>d</i>)	79.3 (<i>d</i>)				20.3 (<i>q</i>)
C(18)	21.1 (<i>q</i>)	17.6 (<i>q</i>)	12.2 (<i>q</i>)				

Fig. 2. Important HMBCs ($\text{H} \rightarrow \text{C}$) and ROESY ($\text{H} \leftrightarrow \text{H}$) correlations of compound **2**

relative β - and α -configuration to $\text{H}-\text{C}(12)$ and $\text{H}-\text{C}(15)$, respectively. The structure of compound **3** was thus established.

The structures of the known compounds were identified by comparison of their physical data with those reported in the literature.

Experimental Part

General. TLC: visualization under UV or by heating at 110° after spraying with 98% $\text{H}_2\text{SO}_4/\text{EtOH}$ 5:95. Column chromatography (CC): silica gel (SiO_2 ; 200–300 mesh; Qingdao Marine Chemical Co., Ltd., P. R. China), Lichroprep RP-18 (40–63 μm ; Merck, Darmstadt, Germany), and Kromasil RP-18 (5 μm , 10×250 mm; Eka Chemicals, Bohus, Sweden). HPLC: Shimadzu instrument (LC-10A pump, SPD-10A UV/VIS detector). Optical rotations: WWZ-2S polarimeter (Shanghai Cany Precision

Instrument Co., Shanghai, P. R. China). UV Spectra: Shimadzu-UV-240 spectrophotometer; λ_{\max} (log ϵ) in nm. IR Spectra: Thermo-Nicolet-Nexus-670 FT-IR spectrometer; KBr pellets; $\tilde{\nu}$ in cm^{-1} . 1D- and 2D-NMR Spectra: Bruker-500-Avance-III spectrometer; at 500 (^1H) and 125 MHz (^{13}C); δ in ppm rel. to Me_4Si and solvent signals as internal references, J in Hz. HR-ESI-MS: Bruker-Apex-III mass spectrometer; in m/z (rel. %).

Plant Material. The leaves of *C. cinerascens* were collected in Guangxi Province of P. R. China and were purchased from the Chinese Herb Transaction Center, Anhui Province, P. R. China. The material was identified by Dr. Gang Ren (Zhejiang University, Hangzhou, P. R. China). A voucher specimen (No. CC 080703) is deposited with the School of Biological and Chemical Engineering, Zhejiang University of Science and Technology, P. R. China.

Extraction and Isolation. The air-dried powder of the plant material (7.0 kg) was extracted with 95% EtOH. The EtOH extract was concentrated *in vacuo* to yield a residue (704 g). The residue was dissolved in H_2O and extracted with CHCl_3 . The CHCl_3 extract (261 g) was partitioned between 95% aq. MeOH and petroleum ether (60–90°) to yield the MeOH-soluble fraction (53 g), which was then subjected to CC (SiO_2 ; $\text{CHCl}_3/\text{MeOH}$ 1:0 → 1:1): *Fractions 1–5*. *Fr. 3* (17 g) was applied to CC (*RP-18* (40–63 μm); $\text{MeOH}/\text{H}_2\text{O}$ 4:6 → 10:0), and then applied to CC (SiO_2 ; $\text{CHCl}_3/\text{acetone}$ 10:1 → 8:2): **1** (35 mg), **3** (15 mg), and **4** (26 mg). *Fr. 4* (16 g) was applied to CC (SiO_2 ; $\text{CHCl}_3/\text{acetone}$ 10:1 → 7:3), and then further purified by semi-prep. HPLC ($\text{MeOH}/\text{H}_2\text{O}$ 65:35): **2** (3 mg), **5** (4 mg), **6** (4 mg), and **7** (4 mg).

Cipadesin G (= (1*S*,4*aS*,5*aS*,7*R*,9*S*,9*aS*,10*S*,11*aS*,13*R*)-7-(Acetyloxy)-1-(furan-3-yl)dodecahydro-8,8,9*a*,11*a*,13-pentamethyl-3,12-dioxo-4*a*,10-ethano-4*aH*-pyrano[4,3-*b*][1]benzoxepin-9-acetic Acid Methyl Ester; **1**): White amorphous solid. $[\alpha]_{\text{D}}^{25} = -10.7$ ($c = 0.075$, CHCl_3). IR: 3447, 2952, 1743, 1735, 1370, 1253, 1229, 1090, 1052. ^1H - and ^{13}C -NMR: Tables 1 and 2. HR-ESI-MS: 553.2432 ($[M + \text{Na}]^+$, $\text{C}_{29}\text{H}_{38}\text{NaO}_5$; calc. 553.2414).

Cipadesin H (= (-)-rel-(4*R*,4*aR*,6*S*,7*aS*,8*R*,10*R*,11*S*,11*aS*,12*aR*)-10,11-Bis(acetyloxy)-4-(furan-3-yl)dodecahydro-4*a*,7*a*,9,9-tetramethyl-13-methylene-2,7-dioxo-6,12*a*-methano-4*H*,12*aH*-pyrano[4,3-*b*][1]benzoxocin-8-acetic Acid Methyl Ester; **2**): White amorphous solid. $[\alpha]_{\text{D}}^{25} = -19.5$ ($c = 0.35$, CHCl_3). IR: 3439, 2962, 1746, 1737, 1378, 1248, 1160, 1083, 1048, 1025. ^1H - and ^{13}C -NMR: Tables 1 and 2. HR-ESI-MS: 609.2284 ($[M + \text{Na}]^+$, $\text{C}_{31}\text{H}_{38}\text{NaO}_{11}$; calc. 609.2311).

Cipadesin I (= (-)-rel-(1*R*,4*R*,4*aR*,5*R*,6*R*,7*aS*,8*R*,10*R*,11*S*,11*aS*,12*aR*)-1,5,10,11-Tetrakis(acetyloxy)-4-(furan-3-yl)dodecahydro-4*a*,7*a*,9,9-tetramethyl-13-methylene-2,7-dioxo-6,12*a*-methano-4*H*,12*aH*-pyrano[4,3-*b*][1]benzoxocin-8-acetic Acid Methyl Ester; **3**): White amorphous powder. $[\alpha]_{\text{D}}^{25} = -42.2$ ($c = 0.120$, CHCl_3). IR: 3437, 2961, 1744, 1735, 1433, 1385, 1227, 1176, 1081, 1060, 1029. ^1H - and ^{13}C -NMR: Tables 1 and 2. HR-ESI-MS: 725.2442 ($[M + \text{Na}]^+$, $\text{C}_{35}\text{H}_{42}\text{NaO}_{15}$; calc. 725.2421).

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